

Speed Breeding: innovative approach for present era of crop breeding MalikireddyJasmitha^{*1}and Maraboina Sandhyarani²

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ABSTRACT

The growing human population and changing environmental conditions significant concern for global food security, with the current improvement rate of several important crops which are inadequate to meet future demand. This slow improvement rate is attributed partly to the long generation time of crop plants. An alternate technique to counter this hurdle is the use of speed breeding technology that shortens the breeding cycle and accelerates the crop research through rapid generation advancement. It can be carried out in numerous ways, one of which involves extending the duration of plant's daily exposure to light, combined with early seed harvest, to cycle quickly from seed to seed, thereby reducing the generation time for some long-day or day-neutral crops. It can also achieve up to 6 generations per year in growth chambers and normal glasshouse conditions. The use of supplemental lighting using LEDs in a glasshouse environment allows rapid generation cycling through single seed descent (SSD) method and plant density can be scaled-up for large crop improvement programs (Watson et al., 2018).

INTRODUCTION

Speed breeding can accelerate backcrossing and pyramiding of traits as well as transgenic pipelines (Chiurugwi et al., 2018). The idea for growing of wheat in space inspired by senior research fellow Dr Lee Hickey of the University of Queensland, Australia. The first spring wheat variety developed using speed breeding is 'DS Faraday', which was released in 2017 in Australia. Speed breeding carries in an enclosed chamber with artificially provided



LED light (e.g., halogen lamps) which is provided PAR of 400-700 nm and photoperiod of 22 hours with 2 hours of darkness in 24 hours of the diurnal cycle and temperature should be maintained accordingly and relative humidity 60-70% also maintained during the entire life cycle (Hickey et al., 2018). Vegetable crops like radish, pea tomato (Introgression of continuous light tolerance gene CAB-13 to increase productivity under continuous light), Amaranthus, Cassava, Potato brassica, Sugar beet and some other leafy vegetables crops (Chiurugwi et al., 2018). Speed breeding is likely to reduce generation time for other crops like tomato, potato, Amaranthus (can produced eight- generations per year instead of two in the field) Pepper (early flowering and fruiting under continuous light) (Kai et al., 2019).

PRINCIPLE

The principle behind speed breeding is to use optimum light intensity (photoperiod) light quality, optimum temperature (varied accordingly crop to crop), and daylight length control (22 h light, 22oC at day/17oC at night with high light intensity) (Ghosh et al., 2018) which leads to accelerate the rate of photosynthesis, stimulates early flowering, seed maturity, harvesting and ultimately shorten the generation time required for crop growth and development. Using speed breeding coupled with single seed descent is more commonly used for developing efficient breeding line and elite inbred line which can be exploited for the development of improved crop varieties in a shorter time, which is cheaper compared to the production of Di-haploids. Speed breeding is also used for gene insertion (common haplotypes) of distinct phenotypes followed by MAS of elite hybrid lines (Alahmad et al., 2018)

Standard protocol, greenhouse design and set up-requirements for speed breeding

To make successful approaches of speed breeding program a proper channel and setup requirements are essential. Controlled green-house structure, the light requirements (Photoperiod), temperature, humidity and germplasm are necessary (Ahmar et al., 2020). Photoperiod (Light) requirement differs in different plant species because different plants respond differently with varying wave-length emitted by different lighting sources. Generally, light requirements covering PAR (400-700 nm) is more suitable for speed breeding with 22 hours of photoperiod. The optimum temperature should be maintained



according to different plant species with specific growth stage (seedling germination, vegetative growth stage, flowering, fruiting, and fruit setting) with 60-70 % relative humidity (Chiurugwi et al., 2018). The following methods are employed for speed breeding program:

(A) Controlled environmental chamber speed breeding condition

This type of structure programmed is run up to 22-hour of photoperiod, with an optimum temperature of 22oC during the photoperiod, and about 17oC during the 2-hour dark period. An artificial photoperiod can be applied by a mixture of white LED bars (Valoya; 6 units per 3.67 m2), far- red LED lamps (Valoya; 12 units per 3.67 m2) and metal HQI lamps (Valoya; 32 units per 3.67 m2) and optimal light intensity should be adjusted to 360- 380 µmol m-2 s -1 (highest value after ramping) at bench height, where pots is planted on the bench. Generally, wheat, barley, amaranthus and pea are suited to model plant for speed breeding under controlled environmental conditions.

(B) Glasshouse speed breeding conditions

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A temperature-controlled greenhouse is fitted and maintained at 17/22°C optimum temperature regime, with sodium vapour lamps and 12 hours of turnover and 22-hour of photoperiod (Ghosh et al., 2018). Light intensity is maintained as 440-650 µmol m-2 s -1 at adult plant height (approximately 45 cm above the bench height).

(C) Homemade growth room design for low-cost speed breeding

A homemade structure about 3 m x 3 m x 3 m with insulated sandwich panelling and fitted lighting equipment about 7 LB-8 LED lightboxes (1 lightbox per 0.65 m2) from Grow Candy (www.growcandy.com). Light quantity and quality about PAR at bench height ranged from 210-260 μ mol m-2 s -1 and at 50 cm above the pot from 340-590 μ mol m-2 s -1. The lights were situated at a height of 140 cm above the bench. In this type of structure, the room can accommodate 90 pots of 8" diameter and 5 L volume are maintained at this kind of structure.

Application and constraint of speed breeding in vegetable improvement

Speed breeding can accelerate the rapid generation and develop the breeding population. Mobini et al., (2016) recorded and reported rapid development of mapping population in pea, mutation studied, fast-forwarding genetic trait, faster and better phenotyping. Watson et al., (2018) recorded that the phenotypes associated with the EMS-



induced mutation of the awn suppressor B1 locus 9 and the green revolution reduced height gene (Rht) in wheat could be accurately recapitulated, encapsulated in the controlled greenhouse condition. Marker- assisted selection coupled with speed breeding provides a flexible platform to rapid introgression of the desired gene for targeted trait having resistance to biotic and abiotic stress and pyramiding could help the development of multiple disease resistance cultivars in different cereals and vegetable crops.

SPEED BREEDING HAS SOME FOLLOWING LIMITATIONS

(A) Most vegetables are long day and day-neutral plants hence, continuous light can accelerate the genetic gain and breeding cycle but, short day plant required limited photoperiod for their growth and development which limits the application of speed breeding for improvement short day vegetable crops. Speed breeding is more straight forward and easier implementation in long day and day-neutral types than short-day plant, but Lee Hickey et al., (2018) have been standardized speed breeding protocol for some short-day plants like millet, sorghum etc, but not in any vegetable crop yet been developed.

(**B**) Different phenotyping: speed breeding accelerates rapid cycling and shorten generation time further early harvest of immature seed can interfere with phenotyping of some trait (Hickey et al., 2009).

(C) High initial investment: speed breeding program are carried in enclosed glass house chambers which requires highly initial investment, purchasing lighting equipment and controlled capabilities and supplementary LED lighting provides more efficient power usage and reduced heat than other lighting types, such as SVLs. (D) Plants which are grown in an enclosed glass chamber with extended photoperiod show various physiological and toxicity symptoms like, chlorosis, necrosis and yellowing due to any micronutrient or heavy metal deficiency or excess.

SUMMARY

Recent advancements in genetic tools, breeding methods and development of NGS sequencing technology provides a flexible platform for sequencing, tagging, mapping and introgression of a gene for the desired trait at low cost, but it is time-consuming. Speed





breeding coupled with other approaches tends to shorten the breeding cycle, generation advancement and accelerates the development of cultivars in a very short time. speed breeding and integration with other breeding technologies may lead rapid genetic gain, for example, tomato is sensitive to constant light, but researchers have identified a tomato gene (CAB-13) that make the plant to tolerate constant long photoperiod when transferred into a tomato cultivar grown under speed breeding conditions (20% increase in fruit yield). Speed breeding as an alternative solution which can enable breeding in other recalcitrant crops, such as short-day species like maize and biennial species like sugar beet. It can be concluded that speed breeding combining other genetic tools like genome editing, Marker assisted backcrossing, Transgenic approaches coupled with cost-effective genotyping and rapid phenotyping helps to develop homozygous inbred line followed by crossing will facilitate rapid cycling, genetic gain and rapid development of improved cultivars to researchers or Plant breeder. Therefore, speed breeding integrated with other multiple branches can be used to improve Orphans, legumes, vegetables, leafy vegetables and other forage crops.

REFERENCES

- Ahmar, S, Rafaqat, A. G Jung K. H, Faheem, A, Qasim, M.U, Zhou, W. 2020. Conventional and molecular techniques from simple breeding to speed breeding in crop Plants: Recent Advances and Future Outlook. Int. J. Mol. Sci. 21(7): 2590.
- Alahmad, S, Dinglasan, E, Leung, K. M. 2018. Speed breeding for multiple quantitative traits in durum wheat. Plant Methods. 14:36.
- Chiurugwi, T, Kemp, S, Powell, W. and Hickey, L. T. 2019. Speed breeding orphan crops. Theoretical and Applied Genetics. 132(3): 607-616.
- Hickey, L. T, Hafeez, A. N, Robinson, H, Jackson, S. A, Leal-Bertioli, S. C, Tester, M, Gao, C, Godwin, I. D, Hayes, B. J. and Wulff, B. B. 2019. Breeding crops to feed 10 billion. Nature biotechnology. 37(7): 744-754.
- Mobini, S. H, Lulsdorf, M, Warkentin, T. D. and Vandenberg, A. 2015. Plant growth regulators improve in vitro flowering and rapid generation advancement in lentil and faba bean. In vitro cellular and Development Biology –Plant. (51): 71-79.
- Voss-Fels, K. P, Stahl, A. and Hickey, L. T., 2019. Q&A: Modern crop breeding for future food security. BMC biology. 17(1): 1-7.